

ACE Taq-Plus DNA Polymerase

Cat# EP1105 – 250U | EP1106 – 1000U | EP1107 – 3000U

Storage: All components should be stored at -20°C.

INTRODUCTION

Taq Plus DNA polymerase is a mixture of Taq DNA polymerase and an enzyme containing 3'→5' exonuclease activity. The fidelity of Taq Plus is 6 times greater than that of Taq DNA Polymerase. Compared with Taq DNA Polymerase, Taq Plus DNA polymerase has stronger amplification performance, higher sensitivity and yield, and is more tolerant of impurities within 5 kb amplifying range. The obtained PCR products are compatible with CE One Step Cloning Kit series (ACE, Cat.No. #EC1001, #EC1008). The PCR products contain A at the 3'-end and can be directly cloned into T-Vectors.

CONTENTS

Component	EP1105 250 U	EP1106 1000U	EP1107 3000U
10× Taq Plus Buffer (Mg ²⁺ plus)	1ml	4ml	4ml*3
Taq Plus DNA Polymerase (5 U/μl)	50ul	200ul	200ul*3

UNIT DEFINITION

One unit (U) is defined as the amount of enzyme that incorporates 10 nmol of dNTPs into acid-insoluble products in 30 min at 74 °C, with activated salmon sperm DNA as the template / primer.

QUALITY CONTROL

Exonuclease Activity: The product is tested in a reaction containing 10 U of Taq DNA Polymerase and 0.6μg of λ-Hind III. After incubation at 37°C for 16 hours, there is no visually discernible change to DNA bands determined by agarose gel electrophoresis.

Endonuclease Activity: The product is tested in a reaction containing 10 U of Taq DNA Polymerase and 0.6μg of Supercoiled pBR322. After incubation at 37°C for 4 hours, there is no visually discernible change to DNA band determined by agarose gel electrophoresis.

Functional Assay: The human α-1-antitrypsin gene is amplified for 30 cycles in a 50μl system using 1.25U of Taq DNA Polymerase and 100 ng human genomic DNA as template. A single DNA band of 360 bp is detected by 1% agarose gel electrophoresis.

PROTOCOL

1. General reaction mixture for PCR:

ddH ₂ O	to 50 μ l
10 \times Taq Plus Buffer (Mg ²⁺ plus)	5 μ l
25 mM MgCl ₂ ^a	Optional
dNTP Mix (10 mM each)	1 μ l
Template DNA ^b	optional
Primer1(10 μ M)	2 μ l
Primer2(10 μ M)	2 μ l
Taq Plus DNA Polymerase(5U/ μ l) ^c	0.5 μ l

a. The final concentration of Mg²⁺ of this mixture is 2 mM, as for most PCR reactions, the optimized final concentration of Mg²⁺ is 1.5 mM- 2 mM. However, if necessary, the concentration of Mg²⁺ can be increased by adding 25 mM MgCl₂.

b. The recommended amount of DNA template for a 50 μ l reaction system is as follows:

Human Genomic DNA	0.1 - 1 μ g
Bacterial Genomic DNA	10 - 100 ng
λ DNA	0.5 - 5 ng
Plasmid DNA	0.1 - 10 ng

c. The amount of Taq DNA Polymerase can be adjusted between 0.25 μ l and 1 μ l. Generally, higher level of Taq will increase the yield of PCR products but may decrease the specificity of PCR amplification.

2. Thermocycling conditions for a routine PCR:

94 $^{\circ}$ C	5 min (Pre-denaturation)	} 30 - 35 cycles
94 $^{\circ}$ C	30 sec	
55 $^{\circ}$ C*	30 sec	
72 $^{\circ}$ C	60 sec / kb	
72 $^{\circ}$ C	7 min (Final extension)	

*The optimal annealing temperature should be 1-2 $^{\circ}$ C lower than the T_m of the primers used.

HANDLING NOTES

Taq DNA Polymerase also shows polymerase activity at room temperature. Therefore, it is recommended to set up reaction systems on ice and then immediately start the reaction in a PCR amplifier, so as to reduce nonspecific amplification during preparation and get better PCR results.

PRIMERS DESIGNING NOTES

1. Choose C or G as the last base of the 3'-end of the primer;
2. Avoid continuous mismatching at the last 8 bases of the 3'-end of the primer;
3. Avoid hairpin structure at the 3'-end of the primer;
4. T_m of the primers should be within the range of 55 $^{\circ}$ C - 65 $^{\circ}$ C;

5. Additional sequence should not be included when calculating T_m of the primers;
6. GC content of the primers should be within the range of 40% - 60%;
7. T_m and GC content of forward and reverse primers should be as similar as possible.

PRODUCT USE LIMITATION

These products are intended for research use only.