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Mouse VEGFA(Vascular Endothelial Growth Factor A) ELISA Kit

Cat# E5044
store at at -20°C or -80°C

INFORMATION

Size	96T
Sensitivity	5.7 pg/mL
Reactivity	Mouse
Detection range	15.63-1000 pg/mL
Specificity	This assay has high sensitivity and excellent specificity for detection of Mouse VEGFA.No significant cross-reactivity or interference between Mouse VEGFA and analogues was observed.Please refer to the outer packaging label of the kit for the specific shelf life.
Special Explanation	1. Store kit at 4°C immediately upon receipt. 2. Do not use the kit after the expiration date. 3. Please check whether all components are complete after opening the package.All kit components have been formulated and quality control tested to function successfully as a kit. Donot mix or substitute reagents or materials from other kit, performance cannot be guaranteed if utilized separately or substituted.
Materials Required, Not Supplied	1. Microplate reader capable of measuring absorbance at 450 ± 10 nm. 2. High-speed centrifuge. 3. Electro-heating standing-temperature cultivator. 4. Absorbent paper. 5. Distilled or deionized water. 6. Single or multi-channel pipettes with high precision and disposable tips. 7. Precision pipettes to deliver 2 µL to 1 mL volumes.
Safety notes	1. This kit is sold for lab research and development use only and not for use in humans or animals. 2. Reagents should be treated as hazardous substances and should be handled with care and disposed of properly. 3. Gloves, lab coat, and protective eyewear should always be worn, Avoid any skin and eye contact with Stop Solution and TMB. In case of contact, wash thoroughly with water.
Test Principle	The test principle applied in this kit is Sandwich enzyme immunoassay. The microtiter plate provided in this kit has been pre-coated with an antibody specific to Vascular Endothelial Growth Factor A(VEGFA). Standards or samples are added to the appropriate microtiter plate wells then with a biotin-conjugated antibody specific to Vascular Endothelial Growth Factor A(VEGFA). Next, Avidin conjugated to Horseradish Peroxidase (HRP) is added to each microplate well and incubated. After TMB substrate solution is added, only those wells that contain Vascular Endothelial Growth Factor A(VEGFA), biotin-conjugated antibody and enzyme-conjugated Avidin will exhibit a change in color. The enzyme-substrate reaction is terminated by the addition of sulphuric acid solution and the color change is measured

	spectrophotometrically at a wavelength of 450nm ± 10nm. The concentration of Vascular Endothelial Growth Factor A(VEGFA) in the samples is then determined by comparing the OD of the samples to the standard curve.
Sample collection and storage	**Serum** - Use a serum separator tube and allow samples to clot for two hours at room temperature or overnight at 4°C before centrifugation for 20 minutes at approximately 1000×g. Assay freshly prepared serum immediately or store samples in aliquot at -20°C or -80°C for later use .Avoid repeated freeze/thaw cycles **Plasma** - Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge samples for 15 minutes at 1000×g at 2-8°C within 30 minutes of collection. Remove plasma and assay immediately or store samples in aliquot at -20°C or -80°C for later use .Avoid repeated freeze/thaw cycles. Tissue homogenates - The preparation of tissue homogenates will vary depending upon tissue type. 1. Tissues were rinsed in ice-cold PBS to remove excess blood thoroughly and weighed before homogenization. 2. Minced the tissues to small pieces and homogenized them in fresh lysis buffer (different lysis buffer needs to be chosen based on subcellular location of the target protein) (w:v = 1:9, e.g. 900 µL lysis buffer is added in 100 mg tissue sample) with a glass homogenizer on ice (Micro Tissue Grinders woks, too). 3. The resulting suspension was sonicated with an ultrasonic cell disrupter till the solution is clarified. 4. Then, the homogenates were centrifuged for 5 minutes at 10000×g. Collection the supernatant and assay immediately or aliquot and store at ≤-20°C. **Cell Lysates** - Cells need to be lysed before assaying according to the following directions. 1. Adherent cells should be washed by cold PBS gently, and then detached with trypsin, and collected by centrifugation at 1000×g for 5 minutes (suspension cells can be collected by centrifugation directly). 2. Wash cells three times in cold PBS. 3. Cells were then resuspended in fresh lysis buffer with concentration of 107 cells/mL. If it is necessary, the cells could be subjected to ultrasonication till the solution is clarified. 4. Centrifuge at 1500×g for 10 minutes at 2-8°C to remove cellular debris. Assay immediately or aliquot and store at ≤-20°C. **Urine** -Aseptically collect the first urine of the day (mid-stream), voided directly into a sterile container. Centrifuge to remove particulate matter, assay immediately or aliquot and store at ≤-20°C. **Saliva** - Collect saliva using a collection device or equivalent. Centrifuge samples for 15 minutes at 1,000×g at 2-8°C. Remove particulates and assay immediately or store samples in aliquot at ≤-20°C. Avoid repeated freeze/thaw cycles. **Cell culture supernatants and other biological fluids** - Centrifuge samples for 20 minutes at 1000×g. Collect the supernatant and assay immediately or store samples in aliquot at -20°C or -80°C for later use.Avoid repeated freeze/thaw cycles.
Note	1. Samples to be used within 5 days may be stored at 4°C, otherwise samples

	must be stored at -20°C (≤1 month) or -80°C (≤2 months) to avoid loss of bioactivity and contamination. Avoid repeated freeze/thaw cycles. 2. Sample hemolysis will influence the result, so hemolytic specimen should not be used. 3. When performing the assay, bring samples to room temperature. 4. If the concentration of the test material in your sample is higher than that of the standard product, please make the appropriate multiple dilution according to the actual situation (it is recommended to do preliminary experiment to determine the dilution ratio.
Summary	1.After the kit is equilibrated at room temperature, add 100µL of standard working Buffer (gradually diluted according to the instructions) or 100µL of sample to each well, incubate at 37°C for 80 minutes. 2. Discard the liquid in the plate, add 200µL of Wash Buffer to each well, and wash the plate 3 times. After spin-drying, add 100µL Biotinylated Antibody working solution to each well,incubate at 37°C for 50 minutes. 3. Discard the liquid in the plate, add 200µL Wash Buffer to each well, and wash the plate 3 times. After drying, add 100µL Streptavdin-HRP working solution to each well, incubate at 37°C for 50 minutes. 4. Discard the liquid in the plate, add 200µL Wash Buffer to each well, and wash the plate 5 times. After spin-drying, add 90µL TMB to each well,incubate at 37°C for 20min.
Reagent preparation	1. Bring all kit components and samples to room temperature (18-25°C) before use. 2. If the kit will not be used up in one time, please only take out strips and reagents for present experiment, and save the remaining strips and reagents as specified. 3. Dilute the 25x wash buffer into 1x working concentration with double steaming water. 4. Standard working solution-Reconstitute the Standard with 1.0mL of Standard Diluent, kept for 10 minutes at room temperature, shake gently(not to foam). The concentration of the standard in the stock solution is 1000 pg/mL. Please prepare 7 tubes containing 0.5mL Standard Diluent and use the diluted standard to produce a double dilution series according to the picture shown below. Mix each tube thoroughly before the next transfer. Set up 7 points of diluted standard such as 1000 pg/mL,500 pg/mL, 250 pg/mL, 125 pg/mL, 62.5 pg/mL, 31.25 pg/mL, 15.63 pg/mL, and the last EP tubes with Standard Diluent is the blank as 0 pg/mL. In order to guarantee the experimental results validity, please use the new standard solution for each experiment. 5. Biotinylated Antibody and Streptavidin-HRP: Briefly spin or centrifuge the stock Biotinylated Antibody and Streptavidin-HRP before use. Dilute them to the working concentration 100-fold with Biotinylated Antibody Diluent and HRP Diluent, respectively. 6. TMB substrate - Aspirate the needed dosage of the solution with sterilized tips and do not dump the residual solution into the vial again.
Note	1. Disposable pipette tips, flasks or glassware are preferred, reusable glassware must be washed and thoroughly rinsed of all detergents before use. 2. Bacterial or fungal contamination of either samples or reagents or

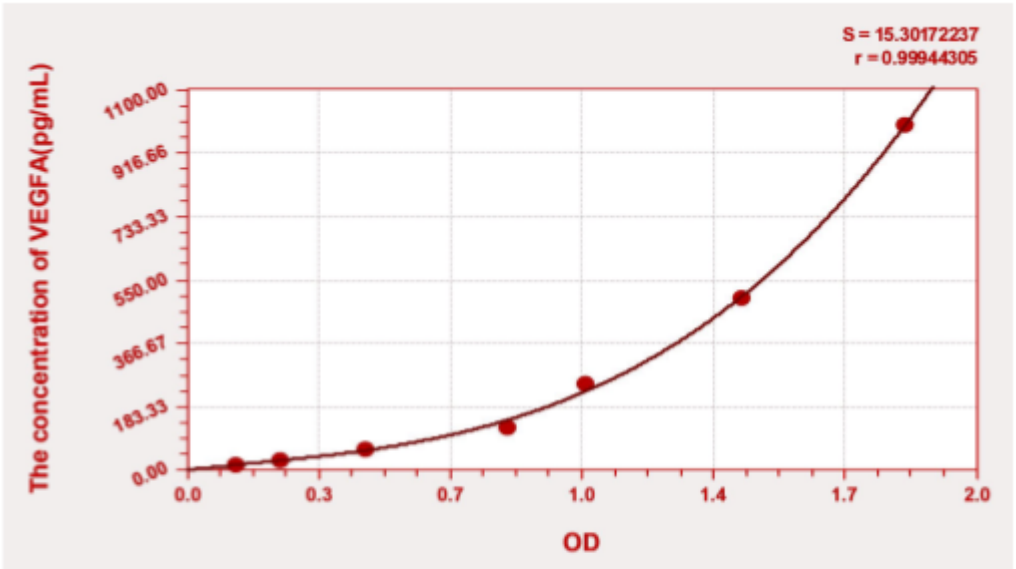
	cross-contamination between reagents may cause erroneous results. 3. All residual washing liquid must be drained from the wells by efficient aspiration or by decantation followed by tapping the plate forcefully on absorbent paper. Never insert absorbent paper directly into the wells. 4. If crystals have formed in the Wash Solution concentrate (25×), warm to room temperature and mix gently until the crystals are completely dissolved. 5. Prepare standards within 15 minutes before assay. This standard can only be used once. 6. The TMB solution is light sensitive. Avoid prolonged exposure to light. Also, avoid contact of the TMB solution with metal to prevent color development. Warning TMB is toxic avoid direct contact with hands. Dispose of properly. If a dark blue color develops within a few minutes after preparation, this indicates that the TMB solution has been contaminated and must be discarded. 7. When pipetting reagents, maintain a consistent order of addition from well-to-well. This will ensure equal incubation times for all wells. Dispense the TMB solution within 15 minutes following the washing of the microtiter plate. 8. It is highly recommended to use the remaining reagents within 1 month provided this is prior to the expiration date of the kit. For the expiration date of the kit, please refer to the label on the kit box.
Samples preparation	1. Equilibrate all materials and prepared reagents to room temperature prior to use. Prior to use, mix all reagents thoroughly taking care not to create any foam within the vials. 2. The user should calculate the possible amount of the samples used in the whole test. Please reserve sufficient samples in advance. 3. Please predict the concentration before assaying. If values for these are not within the range of the standard curve, users must determine the optimal sample dilutions for their particular experiments.
Assay Procedure	1. Determine wells for diluted standard, blank and sample. Prepare 7 wells for standard, 1 well for blank. Add 100 µL each of standard working solution (read Reagent Preparation), or 100 µL of samples into the appropriate wells. Cover with the Plate sealer. Incubate for 80 minutes at 37°C. 2. Remove the liquid of each well. Aspirate the solution and wash with 200 µL of 1× Wash Solution to each well and let it sit for 1-2 minutes. Remove the remaining liquid from all wells completely by snapping the plate onto absorbent paper. Totally wash 3 times. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against absorbent paper. 3. Add 100 µL of Biotinylated Antibody working solution to each well, cover the wells with the plate sealer and incubate for 50 minutes at 37°C. 4. Repeat the aspiration,wash process for total 3 times as conducted in step 2. 5. Add 100 µL of Streptavidin-HRP working solution to each well, cover the wells with the plate sealer and incubate for 50 minutes at 37°C. 6. Repeat the aspiration,wash process for total 5 times as conducted in step 2. 7. Add 90 µL of TMB Substrate Solution to each well. Cover with a new Plate sealer. Incubate for 20 minutes at 37°C (Don't exceed 30 minutes). Protect from light. The liquid will turn blue by the addition of TMB Substrate Solution. 8. Add 50 µL of Stop reagent to each well. The liquid will turn yellow by the addition of Stop reagent.

	Mix the liquid by tapping the side of the plate. If color change does not appear uniform, gently tap the plate to ensure thorough mixing. The insertion order of the Stop reagent should be the same as that of the TMB Substrate Solution. 9. Remove any drop of water and fingerprint on the bottom of the plate and confirm there is no bubble on the surface of the liquid. Then, run the microplate reader and conduct measurement at 450 nm immediately.
Precision	Intra-assay Precision (Precision within an assay) : CV%<8% Three samples of known concentration were tested twenty times on one plate to assess intra-assay precision. Inter-assay Precision (Precision between assays) : CV%<10% Three samples of known concentration were tested in forty separate assays to assess inter-assay precision.

Calculation of Results

Average the duplicate readings for each standard, control, and samples and subtract the average zero standard optical density. Construct a standard curve with the Mouse VEGFA concentration on the y-axis and absorbance on the x-axis,and draw a best fit curve through the points on the graph. If sampleshave been diluted, the concentration read from the standard curve must be multiplied by the dilutionfactor. Using some plot software, for instance, curve expert.

Concentration (pg/mL)	OD	Corrected OD
1000	1.946	1.842
500	1.528	1.424
250	1.128	1.024
125	0.928	0.824
62.5	0.563	0.459
31.25	0.342	0.238
15.63	0.228	0.124
0	0.104	0.000



PRODUCT USE LIMITATION

Recovery

Matrices listed below were spiked with certain level of recombinant VEGFA and the recovery rates were calculated by comparing the measured value to the expected amount of VEGFA in samples.

Matrix	Recovery range	Average
serum(n=5)	85-99%	92%
EDTA plasma(n=5)	96-107%	102%
heparin plasma(n=5)	83-95%	89%

Linearity

The linearity of the kit was assayed by testing samples spiked with appropriate concentration of VEGFA and their serial dilutions. The results were demonstrated by the percentage of calculated concentration to the expected.

Sample	1:2	1:4	1:8	1:16
serum(n=5)	89-103%	79-92%	88-97%	85-97%
EDTA plasma(n=5)	95-102%	97-105%	87-94%	93-101%
Heparin plasma(n=5)	87-95%	78-91%	97-103%	95-101%

Trouble shooting

Problem	Reason	Solution
Poor standard curve	Inaccurate pipetting	Check pipettes
	Improper standard dilution	Prior to opening, briefly spin the stock standard tube and dissolve the powder thoroughly by gentle mixing
	Incomplete washing and aspiration	Adequate washing and adequate aspiration
Low single	Inadequate reagent volumes added to wells	Calibrate pipettes and add adequate reagents
	The samples storage too long	Use new sample and repeat assay
	Incorrect incubation temperature	Ensure the incubator temperature is always 37°C
Poor Precision	Inaccurate pipetting	Check and calibrate pipettes
	Incomplete washing of wells	Ensure sufficient washing
	Contaminated wash buffer	Prepare fresh wash buffer
Poor repeatability	Time of addition is not consistent	Ensure that the sample time is consistent every time
	The washing conditions are not consistent	Ensure the frequency and strength of each wash are consistent