

# ACE Super-Fidelity DNA PCR kit

Cat# EP1005 – 100U / EP1006 - 500U

Storage at -20 °C for one year

## INTRODUCTION

**ACE Super-Fidelity DNA Polymerase** is a new generation superior enzyme based on Pfu DNA Polymerase for robust PCR with extreme fidelity. High amplification efficiency and template adaptability make ACE Super-Fidelity DNA Polymerase suitable for almost all PCR reactions.

The unique extension factor, specificity-promoting factors and plateau-inhibiting factor in ACE Super-Fidelity DNA Polymerase greatly improve its long-fragment amplification ability, specificity, and yield.

ACE Super-Fidelity DNA Polymerase is capable of amplifying long fragments such as 40 kb  $\lambda$ DNA, 20 kb genomic DNA and 10 kb cDNA. The amplification error rate of ACE Super-Fidelity DNA Polymerase is 53-fold lower than that of Taq and 6-fold lower than that of Pfu. In addition, ACE Super-Fidelity DNA Polymerase has a good resistance to PCR inhibitors and can be used for direct PCR amplifications with bacteria, fungi, plants, animal tissues, and even whole blood samples. ACE Super-Fidelity DNA Polymerase contains two monoclonal antibodies inhibiting the 5'  $\rightarrow$  3' polymerase activity and 3'  $\rightarrow$  5' exonuclease activity at room temperature. Super fidelity and supreme amplification efficiency make ACE Super-Fidelity DNA Polymerase the best choice for high fidelity PCR. Amplification will generate blunt-ended products, which are compatible with other cloning Kit

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No	Component	EP1005 – 100 U	EP1006– 500 U
DA	ACE Super-Fidelity DNA Polymerase (1 U/ul)	100 $\mu$ l	100 $\mu$ l x 5
DB	2X ACE Super-Fidelity Buffer	1.25 ml x 2	1.25 ml x 10
DC	10 mM dNTP Mix	100 $\mu$ l	100 $\mu$ l x 5
DD	10X Loading buffer	1.25 ml	1.25 ml x 5

## UNIT DEFINITION

One unit (U) is defined as the amount of enzyme that incorporates 10 nmol of dNTPs into acid-insoluble products in 30 minutes at 74°C. With activated salmon sperm DNA as the template/primer.

## **PROTOCOL**

### **1. Mix the following components**

ACE Super-Fidelity DNA Polymerase (1 U/ul) <sup>a</sup>	1 ul
2X ACE Super-Fidelity Buffer <sup>b</sup>	25 ul
10 mM dNTP Mix	1 ul
Primer 1 (10 uM)	2 ul
Primer 2 (10 uM)	2 ul
Template DNA / cDNA <sup>c</sup>	Optional
ddH <sub>2</sub> O	To 50 ul

**a.** The recommended final concentration of ACE Super-Fidelity DNA Polymerase is 1 U/50  $\mu$ l, which can be optimized between 0.5 U/50  $\mu$ l and 2 U/50 $\mu$ l, if necessary.

**b.** 2X ACE Super-Fidelity Buffer contains Mg<sup>2+</sup>. The final concentration of Mg<sup>2+</sup> is 2 mM.

**c.** Optimal reaction concentration varies from templates. In a 50  $\mu$ l system, there commended template usage is as follows: genomic DNA, 50 ng-400 ng; plasmid or virus DNA, 10 pg-30 ng; cDNA, 1  $\mu$ l-5  $\mu$ l.

### **2. Place the sample in a PCR instrument and run the following program for PCR:**

Stage	Temp.	Time	Cycle
Pre-Denaturation <sup>a</sup>	95°C	30 s / 3 min	1
Denaturation	95°C	15 s	25-35
Annealing <sup>b</sup>	56-72°C	15 s	
Extension <sup>c</sup>	72 °C	30-60s/kb	
Final Extension	95°C	15 s	1

**a.** For pre-denaturation, the recommended time is 30 sec for plasmid/virus DNA and 3 min for genomic DNA / cDNA.

**b.** For annealing, the recommended temperature is the T<sub>m</sub> of the primers. If the T<sub>m</sub> of the primers is higher than 72°C, the annealing step can be removed (two-step PCR). If necessary, the annealing temperature can be further optimized in a gradient. In addition, the amplification specificity depends directly on the annealing temperature. Raising annealing temperature in 3°C increments is helpful to improve poor amplification specificity.

**c.** Longer extension time is helpful to increase the amplification yield.

## For Long-fragment PCR

Use long primers.

When the recommended PCR program does not work, try the Touch Down Two-step PCR as follows:

Stage	Temp.	Time	Cycle
Pre-denaturation	95°C	30 s / 3 min	1
Denaturation	95°C	15 s	5
Extension	74°C	60 s / kb	
Denaturation	95°C	15 s	5
Extension	72°C	60 s / kb	
Denaturation	95°C	15 s	5
Extension	70°C	60 s / kb	
Denaturation	95°C	15 s	25
Extension	68°C	60 s / kb	
Final Extension	68°C	5 min	1

## Primer Design Notes

1. Choose C or G as the last base of the 3'-end of the primer.
2. Avoid continuous mismatching at the last 8 bases of the 3'-end of the primer.
3. Avoid hairpin structure at the 3'-end of the primer.
4. T<sub>m</sub> of the primers should be within the range of 55°C - 65°C.
5. The additional sequence should not be included when calculating T<sub>m</sub> of the primers.
6. GC content of the primers should be within the range of 40% - 60%.
7. T<sub>m</sub> and GC content of forward and reverse primers should be as similar as possible.

## **PRODUCT USE LIMITATION**

These products are intended for research use only.